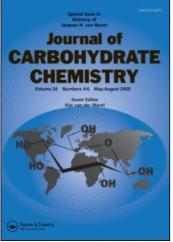
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Journal of Carbohydrate Chemistry

Publication details, including instructions for authors and subscription information: http://www.informaworld.com/smpp/title~content=t713617200

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To cite this Article Boons, G. J. P. H., Steyger, R., Overhand, M., van der Marel, G. A. and van Boom, J. H.(1991) 'Synthesis of Naturally Occurring Ld-Hep*P* Containing Disaccharides', Journal of Carbohydrate Chemistry, 10: 6, 995 – 1007

To link to this Article: DOI: 10.1080/07328309108543968 URL: http://dx.doi.org/10.1080/07328309108543968

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SYNTHESIS OF NATURALLY OCCURRING LD-HEPP

CONTAINING DISACCHARIDES

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March 7, 1991 - Final form July 23, 1991

ABSTRACT

Diastereoselective hydroxymethylation of a suitably protected α -D-manno-hexodialdo-1,5-pyranoside with the Grignard reagent derived from (phenyldimethylsilyl)methyl chloride gives, after additional protecting group manipulations, an easy access to one donor and two acceptors of LD-Hepp. The latter derivatives could be applied successfully for the preparation of the disaccharides α -D-GlcpN-(1-7)-L- α -D-Hepp-OMe and L- α -D-Hepp-(1-6)-L- α -D-Hepp-OMe.

INTRODUCTION

Lipopolysaccharides (LPS) are characteristic cell surface elements¹ of Gram-negative bacteria. The inner core region of LPS consists mainly of L-glycero-D-mannoheptopyranose (LD-Hepp) and 3-deoxy-D-manno-octulosonic acid (KDO), one of which may be linked to Lipid A. Many antibodies (poly or monoclonal) raised against LPS of certain Gram-negative bacteria (particularly rough-mutant bacteria) are believed² to be directed against the common LD-Hepp and KDO containing region of LPS. As part of an ongoing program directed toward the synthesis of immunogenic inner core fragments, we developed suitable methods for the preparation of the non-common sugars LD-Hepp^{3,4} and KDO⁵. With the objective to widen the scope of our LD-Hepp building units, we report here the synthesis of the antigenic disaccharides α -D-GlcpN-(1 \rightarrow 7)-L-

```
L-\alpha-D-Hepp

1

\downarrow

6

\beta-D-Glcp-(1\rightarrow4)-L-\alpha-D-Hepp-(1\rightarrow2)-L-\alpha-D-Hepp

3

\downarrow

1

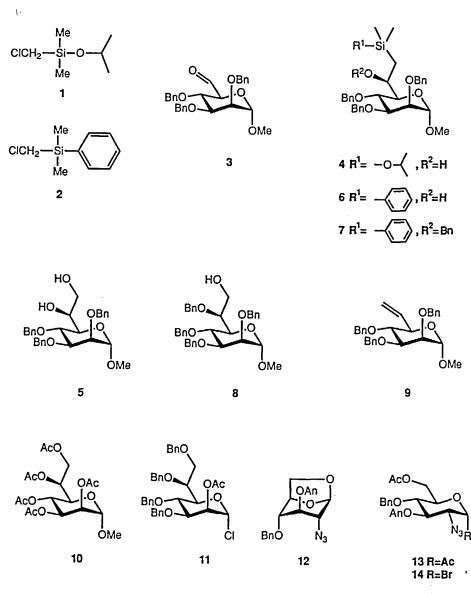
\alpha-D-GlcpN-(1\rightarrow7)-L-D-Hepp
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Figure 1.

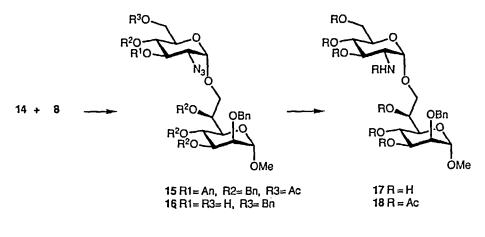
 α -D-Hepp-OMe (*i.e.*, 17) and L- α -D-Hepp-(1 \rightarrow 6)-L- α -D-Hepp-OMe (*i.e.*, 22) which are fragments (see Figure 1) of the inner core LPS structure from Aeromonas hydrophila.⁶

RESULTS AND DISCUSSION

A crucial requirement in a synthetic route to the dimers 17⁷ and 22 is, apart from the stereoselective introduction of the α -interglycosidic linkages, the availability of suitably protected LD-Hepp derivatives. Earlier we reported³ that LD-Hepp building units were accessible by a two-step Tamao procedure.³ For example, condensation of the Grignard reagent derived from (isopropyloxydimethylsilyl)methyl chloride (1) with the fully benzylated α -D-manno-hexodialdo-1,5-pyranoside (3) results in the formation of the silane derivative 4 with a high degree of diastereoselectivity. Unfortunately, it was imperative, due to the instability of the carbon-silicon bond, to convert 4 immediately by oxidation into the stable LD-Hepp derivative 5. Despite this drawback, the usefulness, of the hydroxymethylene extension procedure has been nicely illustrated^{9,10} in the synthesis of several LD-Hepp containing cell wall oligosaccharides. Nonetheless, it has to be noted that the two-step hydroxymethylene extension procedure affords a vicinal diol function which demands in some cases (e.g. introduction of interglycosidic linkages at C-6 or C-7 of LD-Hepp) additional protecting group manipulations.¹⁰ Recently we showed^{4,11} that the stability of the newly introduced silicon-carbon bond could be increased by using (phenyldimethylsilyl)methyl chloride (2) as the hydroxymethylene reagent. Thus,



condensation of the Grignard reagent of 2 with the aldehyde 3 gives in a highly diastereoselective manner the stable silane derivative 6, as evidenced by the following experiments. Cleavage of the carbon-silicon bond¹² in 6 with peracetic acid, in the presence of sodium bromide, gave exclusively the LD-Hepp derivative 5 having the same configuration at C-6 (*i.e.*, L-glycero) as in 5 prepared via the Tamao procedure. Further, benzylation¹³ of 6 followed by the unmasking of the silyl function in 7, under the condition mentioned earlier for the conversion of $6\rightarrow 5$, resulted in a good recovery of 8: thus demonstrating the stabilizing effect of the phenyl group on the newly introduced

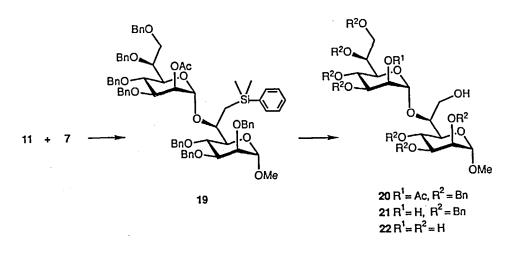




silicon-carbon bond. In this respect, it is also noteworthy to mention that the unmasking of $7\rightarrow 8$ was not accompanied by the formation of the Peterson elimination¹⁴ product 9. Apart from this, three valuable LD-Hepp derivatives emerged from the above executed chemical operations. Thus compounds 8 and 6 may serve as acceptors in glycosidations resulting in the disaccharides 17 and 22, respectively. On the other hand, the LD-Hepp derivative 5 can be readily converted into the LD-Hepp derivative 11, which in turn may serve as donor in a condensation reaction leading to disaccharide 22. Thus hydrogenolysis of 5 followed by acetylation gave 10 which was further elaborated, according to a published¹⁵ procedure, to yield the LD-Hepp donor 11.

At this stage of the synthesis we turned our attention to the preparation of the required disaccharides 17 and 22. In the first instance, the disaccharide 17 containing the D-glucosamine unit $\alpha(1\rightarrow7)$ -linked with LD-Hepp was obtained as outlined in Scheme 1.

Condensation of the LD-Hepp acceptor 8 with glucosyl bromide 14, prepared via acetolysis¹⁶ of the known 1,6-anhydro-3-O-anisoyl-2-deoxy-2-azido-D-glucopyranoside¹⁷ (12) and subsequent treatment of 13 with titanium tetrabromide,¹⁶ was executed under Helferich conditions¹⁸ [Hg(CN)₂/HgBr₂]. Work up, after three days at ambient temperature, and purification gave the homogeneous α -linked disaccharide 15 in 81% yield. Basic hydrolysis of the acyl protecting groups (15 \rightarrow 16) followed by reduction of the azido function and simultaneous hydrogenolysis of the benzyl groups with palladium on charcoal gave the fully deprotected disaccharide 17. Acetylation of 17 gave





exclusively 18, the ¹H- and ¹³C NMR data of which were in excellent agreement with the same disaccharide recently isolated and fully characterized by Kaca *et al.*.¹⁹

In the next stage the feasibility (see Scheme 2) to apply the silane derivative 6 as an acceptor in the condensation with donor 11 was explored. Preliminary experiments indicated that the use of the promoter silver triflate²⁰ was accompanied by Peterson elimination: *i.e.*, conversion of 6 into 9. However, the latter side-reaction could be avoided by using dry and high-quality silver triflate. Thus coupling of 6 with 11 in the presence of silver triflate afforded, after purification, the fully protected disaccharide 19 in 45% yield. Unmasking of the silyl function by oxidation $(19\rightarrow 20)$ followed by Zemplén deacetylation $(20\rightarrow 21)$, and finally hydrogenolysis, resulted in the isolation of homogeneous 22, the ¹H- and ¹³C NMR data of which were in complete accordance with the proposed structure. In conclusion the data presented in this paper firmly support our earlier reported^{4,11} preliminary results. Furthermore, we believe that the commercially available²¹ reagent 2 promises²² to be a very reliable and versatile tool, in terms of diastereoselectivity, protecting group strategy and compatibility with glycosidation conditions, for the preparation of valuable LD-Hepp derivatives.

EXPERIMENTAL

General methods and materials. Dichloromethane, 1,2-dichloroethane and toluene were distilled from P_2O_5 and stored over molecular sieves 4Å. Diethyl ether and

tetrahydrofuran were distilled from CaH₂ and redistilled from LiAIH₄ before use. N,Ndimethylformamide was stirred with CaH₂ for 16 h, then distilled under reduced pressure and stored over molecular sieves 4Å. Methanol was dried by refluxing with magnesium methoxide, distilled and stored over molecular sieves 3Å. TiBr, was sublimed under reduced pressure and stored as a standard solution in dichloromethane. Silver triflate was purchased from Fluka and dried in vacuo, with the exclusion of light. Reactions were performed under strict anhydrous conditions unless noted otherwise. Evaporation was carried out below 40 °C under reduced pressure. Column chromatography was performed on columns of silica gel (Merck, 70-230 mesh). Gel filtration was performed on Sephadex LH-20 (Pharmacia). TLC analyses were conducted on Schleicher and Schüll Fertigfolien F 1500 LS 254. Compounds were visualized by UV light (254 nm) or by spraying with conc. H₂SO₄/methanol (1/10, v/v) and charring at 140 °C. Optical rotations were determined with a Perkin-Elmer Model 241 polarimeter at 20 °C. NMR spectra were recorded with a Jeol JNM-FX200 ('H and "C at 200 and 80.7 MHz, respectively), or a Bruker WM-300 spectrometer equipped with an Aspect-2000 computer (¹H, 300 MHz). Chemical shifts are given in ppm (δ) relative to TMS as internal standard.

Methyl 2,3,4-tri-O-benzyl-7-(phenyldimethyl)silane-7-deoxy-L-glycero- α -D-mannopyranoside (6). A small amount of (phenyldimethylsilyl)methyl chloride

(11.7 g, 60 mmol) in THF (45 mL) was added under a N2 atmosphere to dry magnesium turnings (1.46 g, 60 mmol). The reaction mixture was heated until reflux and, after the reaction was initiated by the addition of 1,2-dibromoethane (0,1 mL), the remaining chloride was added at such a rate as to maintain a gentle reflux. The thus obtained Grignard reagent was transferred to a dry round-bottom flask. The aldehyde 3 was coëvaporated with toluene (2 x 20 mL), dissolved in THF (45 mL) and added dropwise to the cooled (0 °C) Grignard reagent . After stirring for 2 h, TLC analysis (ether/pet. ether, 1/2, v/v) indicated complete conversion of 3 in to 6 (Rf 0.5). The mixture was slowly poured into aqueous ammonium chloride (50 mL, 20%) and extracted with CH₂Cl₂ (200 mL). The organic layer was washed with water (50 mL), dried (MgSO₄) and concentrated under reduced pressure. The oil thus obtained was applied onto a column of silica gel (200 g) and eluted with light pet. ether followed by light pet. ether/ethyl acetate (9/1 \rightarrow 4/1, v/v). Concentration of the appropriate fractions gave 6 as an oil. Yield 13.1 g (71 %), R_f 0.5 (light pet. ether/ether, 2/1, v/v), $[\alpha] + 11.5^{\circ}$ (c 1, CHCl₃). ¹H NMR (CDCl₃): δ 7.57-7.17 (m, 20H, H-arom), 4.91-4.59 (m, 6H, CH₂ Bn), 4.71 (d, 1H, H-1, $J_{12}=2.0$ Hz), 4.05 (m, 2H, H-4, H-6, $J_{43}=J_{34}=9.5$ Hz), 3.84 (dd, 1H, H-3, J₂₃=3.0 Hz), 3.77 (dd, 1H, H-2), 3.34 (1-H, H-5, J₅₅=1.5Hz), 1.37 (dd, 1H, H-7a, $J_{7a,6}=12$ Hz, $J_{7a,7b}=17$ Hz), 0.94 (dd, 1H, H-7b, $J_{6,7b}=5.0$ Hz), 0.36, 0.35 (2xs, 6H,

Si(CH₃)₂); ¹³C NMR (CDCl₃): δ 138.3 (Cq, arom.), 133.5-127.46 (CH arom.), 99.4 (C-1), 80.3, 75.4, 75.1, 74.7, 67.1 (C-2, C-3, C-4, C-5, C-6), 75.3, 72.8, 72.1 (3x CH₂, Bn), 54.7 (OCH₃), 21.7 (C-7), -2.4 ((CH₃)₂Si).

Anal. Calcd for C37H44O6Si: C,72.52; H,7.24. Found: C,72.63; H,7.30.

Methyl 2,3,4,-tri-O-benzyl-L-glycero- α -D-manno-heptopyranoside (5). Acetic acid (15 mL), sodium acetate (2.0 g) and potassium bromide (0.25 g, 2.4 mmol) were added to 6 (0.93 g, 2.0 mmol) and the mixture was stirred until the salts were dissolved. The mixture was cooled (10 °C) and peracetic acid (10 mL, 30% in acetic acid) was added dropwise under the exclusion of light. During the addition gas was liberated. After the mixture was stirred for 1.5 h at 20 °C, TLC analysis (acetone/CH₂Cl₂, 3/97, v/v) indicated complete conversion of the starting material into a more hydrophilic product. The mixture was poured in an aqueous solution of sodium thiosulfate (50 mL, 15 %) and extracted with CH₂Cl₂ (100 mL). The organic layer was washed with water (50 mL), aqueous sodium bicarbonate (50 mL, 10 %), water (50 mL), dried (MgSO₄) and concentrated under reduced pressure. The residue was applied onto a column of silica gel (10 g) and eluted with CH₂Cl₂ followed by CH₂Cl₂/methanol (1/99 \rightarrow 1/49, v/v). Evaporation of the appropriate fractions afforded 5 as an oil. Yield 0.69 g (72 %), Rf 0.3 (methanol/CH₂Cl₂, 3/97, v/v), [α] +30.5° (c 1, CHCl₃).

Methyl 2,3,4,6-tetra-O-benzyl-7-(phenyldimethyl)silane-7-deoxy-L-glycero- α -Dmanno-heptopyranoside (7). N-tetrabutylammonium iodide (0.74 g, 2.0 mmol), sodium hydride (0.77 g, 32 mmol) and benzyl bromide (3.1 mL, 26 mmol) were added to a cooled (0° C) solution of 6 (13.1 g, 21.5 mmol) in DMF (60 mL). After stirring for 3 h at room temperature, TLC analysis (pet. ether/ether, 2/1, v/v) indicated complete conversion of 6 into 7. The reaction mixture was quenched with methanol (10 mL) and concentrated under reduced pressure. The residue was partitioned between CH₂Cl₂ (150 mL) and water (2x50 mL). The organic layer was dried (MgSO₄), and concentrated to an oil, which was applied onto a column of silica gel (200 g) and eluted with light pet. ether followed by light pet. ether /ether (9/1 \rightarrow 4/1, \sqrt{v}). Concentration of the appropriate fractions gave 7 as an oil. Yield 14.3 g (95 %). R_t 0.8 (pet. ether/ether, 2/1, v/v), [α] + 24.6° (c 1, CHCl₃). ¹H NMR (CDCl₃):δ 7.58-7.10 (m, 25H, H-arom.), 4.85 (d, 1H, H-1, $J_{12}=2.0$ Hz), 4.78-4.20 (m, 8H, CH₂, Bn),4.09, t, 1H, H-4, $J_{34}=J_{45}=9.5$ Hz), 4.02 (m,1H, H-6), 3.83 (dd, 1H, H-3, J₂₃=3.0 Hz), 3.76 (dd, 1H, H-2), 3.46 (dd, 1H, H-5, J_{5,6}=1.5 Hz), 3.27 (s, 3H, OCH₃), 1.42 (m, 2H, H-7a, H7b), 0.36, (s, 6H, CH₃Si). ¹³C NMR (CDCl₃): δ 138.8-138.4 (Cq, arom.), 133.6-127.8 (CH, arom.), 99.1 (C-1), 80.4, 75.0, 74.5, 74.4, 72.9 (C-2, C-3, C-4, C-5, C-6), 74.3, 72.3 71.9, 69.8 (4x CH₂, Bn), 55.0 (OCH₃), 17.0 (C-7), -2.1,-2.3 (2x CH₃Si).

Anal. Calcd for C44H50O6Si: C,75.18; H,7.17. Found: C,75.31; H,7.26.

Methyl 2,3,4,6-tetra-O-benzyl-L-glycero-α-D-manno-heptopyranoside (8). Compound 7 (1.4 g, 2.0 mmol) was oxidised with peracetic acid (10 mL) and potassium bromide (0.286 g, 2.4 mmol) in a mixture of acetic acid (15 mL) and sodium acetate (2.0 g), as described for the conversion of 6→5, to give 8 (0.84 g, 72 %), [α] + 37.8° (c 1.1, CHCl₃). ¹H NMR (CDCl₃): δ 7.48-7.18 (20H, arom. H), 4,84 (d, 1H, H-1, J₁₂=2.0 Hz), 4.96-4.39 (m, 8H, CH₂, Bn), 4.21 (t, 1H, H-4, J_{3,4}=J_{4,5}=9.5 Hz), 3.98-3.90 (m, 4H, H-3, H-6, H-7a, H-7b), 3.84 (dd, 1H, H-2, J_{2,3}=3.0 Hz), 3.77 (dd, 1H, H-5, J_{5,6}=1.0 Hz), 3.36 (OCH₃). ¹³C NMR (CDCl₃): δ138.4-138.0 (Cq, arom.), 128.1-127.3 (CH, arom.), 98.8 (C-1), 80.2, 75.7, 74.1, 74.0, 72.5 (C-2, C-3, C-4, C-5, C-6), 74.4, 72.3, 71.9, 71.6 (4x CH₂, Bn), 62.0 (C-7), 54.6 (OCH₃).

Anal. Calcd for C36H40O7: C,73.95; H,6.90. Found: C,74.08; H,6.82.

1,6-di-O-acetyl-3-O-(4-methoxybenzoyl)-4-O-benzyl-2-azido-2-deoxy-β-Dglucopyranose (13). Coumpound **12** (9.4 mmol) was dissolved in a mixture of acetic anhydride (135 mL) and trifluoroacetic acid. After stirring for 24 h, TLC analysis (acetone/CH₂Cl₂, 3/97, v/v) indicated complete conversion of the starting compound into a more hydrophilic product (Rf 0.6). The mixture was diluted with toluene (50 mL) and concentrated under reduced pressure to give, after coevaporation (3x50 mL) of the resulting oil, compound **13** as an amorphous material. Yield 4.1 g (100 %), Rf 0.6 (acetone/CH₂Cl₂, 3/97, v/v). ¹H NMR (CDCl₃): δ 8.07-6.95 (m, 9H, H-arom.), 6.31 (d, 1H, H-1, J₁₂=3.0 Hz), 5.81 (t, 1H, H-3, J₂₃=J₃₄=10 Hz), 4.61-4.41 (AB, 2H, CH₂, Bn), 4.29 (d, 2H, H6a,b), 4.09 (dt, 1H, H-5, J₄₅=10 Hz), 3.78 (t, 1H, H-4), 3.87 (s, 3H, OCH₃), 3.61 (dd, 1H, H-2), 2.19, 2.09 (2xs, 2xCH₃, Ac); ¹³C NMR: δ 170.2, 168.6, 164.9 (3x C=O), 136.6, 121.3 (2xCq, arom.), 131.6, 113.7 (CH, arom), 92.7 (C-1), 74.8 (CH₂, Bn), 75.1, 72.4, 70.9, (C-3, C-4, C-5), 60.8 (C-6), 60.8 (C-2), 55.4 (OCH₃), 20.9, 20.7 (2xCH₃, Ac).

Anal. Calcd for C25H27O9N3: C,58.48; H,5.30. Found: C,58.57; H,5.37.

3-O-(4-methoxybenzoyl)-4-O-benzyl-2-azido-2-deoxy- α -D-glucopyranosyl bromide (14). A solution of TiBr₄ (2.4 mmol, 4 mL) in CH₂Cl₂ (0.23g/mL), was added to a solution of 13 (0.65 g, 1.2 mmol) in CH₂CL₂ (8 mL) and ethyl acetate (0.8 mL). The dark red solution was stirred overnight at 20 °C under an atmosphere of dry nitrogen. TLC analysis (acetone/CH₂Cl₂, 3/97, v/v) showed complete conversion of the starting material into bromide 14 (Rf 0.9). Dry sodium acetate was added until the solution became colourless. The mixture was taken up in dry toluene and filtrated over a pad of Celite and concentrated to dryness, to afford 14 as a colourless oil which was used directly in the next step.

Methyl 2,3,4,6-tetra-O-benzyl-7-O-(3-O-(4-methoxybenzoyl)-4-O-benzyl-6-Oacetyl-2-azido-2-deoxy-α-D-glucopyranosyl)-L-glycero-α-D-manno-heptopyranoside

(15). To a mixture of 8 (0.47 g, 0.8 mmol), Hg[CN]₂ (0.20 g, 0.87 mmol), HgBr₂ (0.025 g, 0.07 mmol) and powdered molecular sieves (0.5 g, 4 Å) in CH₂Cl₂ (4 mL) was added dropwise a solution of the bromide 14 (0.58 g, 1.2 mmol) in CH₂Cl₂. After stirring for 3 days at room temperature, TLC analysis (acetone/CH₂Cl₂, 1/99, v/v) showed the absence of acceptor 8. The mixture was diluted with CH₂Cl₂ (25 mL), filtrated over a pad of Celite and the filtrate was washed with aqueous potassium bromide (2x10 mL, 20%), aqueous sodium bicarbonate (10 mL, 10%) and water (10 mL). The organic layer was dried (MgSO₄), concentrated under reduced pressure and the residue was applied onto a column of silica gel (10 g) and eluted with CH₂Cl₂ followed by acetone/CH₂Cl₂ $(1/99 \rightarrow 1/49, v/v)$. Concentration of the appropriate fractions afforded 15 as an oil. Yield 0.68 g (81 %), $[\alpha]_{e} = 61.1$ (c 1, CHCl₃), Rf 0.5 (acetone/CH₂Cl₂, 1/99, v/v). ¹H NMR (CDCl₃): δ 8.1 (d, 2H, H-2, H-6 An.), 7.43-7.15 (m, 25H, H-Bn), 6.93 (d, 2H, H-3, H-4, An.), 5.86 (dd, 1H, H-3', J_{2'3}=10.5 Hz, J_{3'4}=9.0 Hz), 5.03 (d, 1H, H-1', J_{1'2}=3.5 Hz), 4.83 (d, 1H, H-1, J₁₂=2.0 Hz), 4.93-4.36 (m, 6H, 3x CH₂, Bn), 4.27 (d, 2H, H-6'ab, $J_{3,5}=3.2$ Hz), 4.22 (t, 1H, H-4, $J_{3,4}=J_{4,5}=9.5$ Hz), 4.13 (m, 1H, H-5'), 3.97 (m, 1H, H-6), 3.93 (m, 2H, H-4, H-4'), 3.86 (s, 3H, OCH₃), 3.80 (dd, 2H, H-7ab), 3.61 (m, 1H, H-5), 3.43 (dd, 1H, H-2'), 2.05 (s, 3H, CH₃, Ac.). ¹³C NMR (CDCl₃): δ 169.9 (C=O, Ac.), 164.6, (C=O, An.), 163.3 (C-4, An.), 138.3-136.6 (Cq, Bn), 131.4 (C-2, C-6, An.), 121.3 (C-1, An.), 113.3 (C-3, C-5, An.),99.1, 98.2 (C-1, C-1'), 80.3, 75.9, 75.0, 74.4, 74.1,72.5, 71.7, 68.9 (C-2, C-3, C-4, C-5, C-6, C-3', C-4', C-5'), 74.6, 74.2, 72.8, 72.4, 71.9 (5x CH₂, Bn), 69.0 (C-7), 62.5, (C-6'), 61.6 (C-2'), 54.9, 54.5 (2x OCH₄), 20.3 (CH₄, Ac.).

Anal. Calcd for C₅₉H₆₃O₁₄N₃: C,68.26; H,6.12. Found: C,68.37; H,6.13.

Methyl 2,3,4,6-tetra-O-benzyl-7-(4-O-benzyl-2-azido-2-deoxy- α -D-glucopyranosyl)-L-glycero- α -D-manno-heptopyranoside (16). Compound 15 (0.68 g, 0.66 mmol) was dissolved in a mixture of dioxane, methanol and aqueous NaOH (16 mL) and left for 3 h at 50 °C. TLC analysis (acetone/CH₂Cl₂, 1/19, v/v) indicated complete conversion of the starting material into a more hydrophilic product (R₁ 0.5). The mixture was diluted with water (10 mL) and extracted with CH₂Cl₂ (25 mL). The organic phase was dried (MgSO₄), concentrated under reduced pressure and the residual oily product was applied onto a column of silica gel (10 g) and eluted with CH₂Cl₂ followed by acetone/CH₂Cl₂ (1/49 \rightarrow 1/19, v/v). Concentration of the appropiate fractions afforded 16 as an oil. Yield 0.52 g (92 %), [α] 1.071 (c 1, CHCl₃), Rf 0.5 (acetone/CH₂Cl₂, 1/19, v/v). ¹H NMR (CDCl₃): δ 7.41-7.12 (25H, H-arom.), 5.03 (d, 1H, H-1', J_{1'2}=3.5 Hz), 4.22 (d, 1H, H-1, J₁₂=2.0 Hz), 4.93-4.35 (m, 8H, CH₂, Bn), 4.22-3.42 (m, 13H, H-2 - H7a,b, H-2' -H6'a,b), 3.85 (s, 3H, OCH₃).

Anal. Calcd for C49H55O11N3: C,68.28; H,6.43. Found: C,68.21; H,6.52.

Methyl 2,3,4,6-tetra-O-acetyl-7-O-(2-acetamido-3,4,6-tri-O-acetyl-2-deoxy- α -Dglucopyranosyl)-L-glycero- α -D-manno-heptopyranoside (18). A mixture of 16 (90 mg, 0.21 mmol) and 10 % Pd-C (90 mg) in tert-butanol (5 mL), acetic acid (1 mL) and water (1 mL) was stirred under H₂ for 48 h at 20 °C. The catalyst was removed by filtration and washed with water. The combined filtrates were concentrated under reduced pressure. The remaining residue was concentrated with pyridine (2x10 mL) and dissolved in a mixture of pyridine (5 mL) and acetic anhydride (5 mL). After stirring for 18 h, the mixture was diluted with toluene (10 mL) and concentrated under reduced pressure. The crude oil was dissolved in CH_2Cl_2 (25 mL) and washed with aqueous sodium bicarbonate (10 mL) and water (10 mL). The dried (MgSO₄) organic layer was concentrated under reduced pressure and the resulting material was applied onto a column of silica gel (1 g) and eluted with CH₂Cl₂ followed by CH₂Cl₂/methanol (99/1, v/v). Concentration of the appropriate fractions gave 18 as an oil. Yield 0,12 g (89%), Rf 0.5 (methanol/CH₂Cl₂, 3/97, v/v), [α] 1.071 (c 1, CHCl₃) +62.3° (c 1, CHCl₃). ¹H NMR: δ 6.08 (d, 1H, NH, J_{NH2}=9.2 Hz), 5.68 (t, 1H, H-4, J₃₄=₄₅=9.9 Hz), 5.58 (dd, 1H, H-3, J₂₃=3.3 Hz), 5.50 (m, 2H, H-2, H-3'), 5.42 (ddd, 1H, H-6, J₆₇₁=_{3.6} Hz, J_{6.75}=8.1 Hz, J_{5,6}=2.5 Hz), 5.31 (t, 1H, H-4', J_{3',4}:=J_{4'5}:=9.8 Hz), 4.80 (d, 1H, H-1'), 4.56 (ddd, 1H, H-2', J_{12} =3.8 Hz, J_{23} =10.1 Hz), 4.50 (d, 1H, H-1, J_{12} =1.8 Hz), 4.39 (dd, 1H, H6'a, J_{5',6'a}=5.1 Hz, J_{6'a6'b}=12.3 Hz), 4.16 (dd, 1H, H6'b, J_{5'6'b}=2.6 Hz), 4.00 (ddd, 1H, H-5'), 3.94 (dd, 1H, H-5), 3.80 (dd, 1h, H-7a, J_{7a,b}=11.6 Hz), 3.68 (dd, 1H, H7b), 3.01 (OCH₃), 1.87, 1.81, 1.76, 1.72, 1.71, 1.67 (24H, CH₃, Ac).

Anal. Calcd for C₂₆H₄₃O₁₈N: C,47.49; H,6.59. Found: C,47.57; H,6.63.

Methyl 2,3,4-tri-O-benzyl-6-O-(2-O-acetyl-3,4,6,7-tetra-O-benzyl-L-glycery-Dmanno-heptopyranosyl)-7-deoxy-7-(phenyldimethyl)silane-L-glycero-D-mannoheptopyranoside (19). To a solution of compound 6 (0.165 g, 0.27 mmol) in CH₂Cl₂ (5 mL) was added activated powdered molecular sieves (0.5 g) and the mixture was stirred for 2.5 h in an atmosphere of nitrogen. Silver triflate (0.13 g 0.5 mmol) was then added under the exclusion of light. The mixture was cooled to -30 °C and the chloride 11 (0.30 g, 0.45 mmol) in CH₂Cl₂ (2 mL) was added over a period of 30 min. The reaction mixture was stirred at -20 °C for 16 h. The mixture was filtrated and the filtrate was diluted with CH₂Cl₂ (50 mL) and washed with aqueous sodium thiosulfate (15 mL), sodium bicarbonate (15 mL), water (20 mL), dried (MgSO₄) and concentrated under reduced pressure. The residue was applied to a column of silica gel (2g) and eluted with light pet. ether/ethylacetate (85/15, v/v). Fractions containing the required dimer were concentrated under reduced pressure and the residual oil was applied on a LH-20 Sephadex column (eluent, CH₂Cl₂/methanol). The appropiate fractions were concentrated to give pure 19 in a yield of 0.145 g (45 %) as an oil. Rf 0.3 (acetone/CH₂Cl₂, 3/97, v/v), $[\alpha]_{\circ}$ +18.2° (c 0.95, CHCl₃). ¹³C NMR (CDCl₃): δ 170.7 (C=0, Ac.), 139.6-138.4 (Cq, Arom.), 134.1-127.8 (CH, Arom.), 99.4, 95.3 (C-1, C-1'), 80.6, 76.3, 74.4, 74.3, 72.9, 69.5 (C-2, C-3, C-4, C-5, C-6, C-2', C-3', C-4', C-5', C-6'), 73.8, 73.7, 72.9, 70.8, (CH₂, Bn., C-7'), 55.9 (OCH₃), 21.9 (CH₃, Ac.), 16.8 (C-7), -1.4, -1.8 ((CH₃)₂Si); J_{C1-H1} = 167 Hz, J_{C1-H1} = 171 Hz.

Anal. Calcd for C74H82O13Si: C,73.61; H,6.84. Found: C,73.69; H,6.90.

Methyl 2,3,4-tri-O-benzyl-6-O-(2-O-acetyl-3,4,6,7-tetra-O-benzyl-L-glycero- α -Dmanno-heptapyranosyl)-L-glycero- α -D-manno-heptopyranoside (20). Compound 19 (65 mg, 0.05 mmol) was oxidised, as described earlier for $6\rightarrow$ 5, to give, after further work up and purification homogeneous 20 in a yield of 48 mg (88 %), Rf 0.4 (acetone/ CH₂Cl₂, 3/97, v/v), [α] + 29.9° (c 0.96, CHCl₃). ¹H NMR (CDCl₃): δ 7.32-7.33 (35H, H-arom.), 5,43 - 3.87 (H-1 - H-7_{a,b}, H-1' - H-7'_{a,b}, 7xCH₂, Bn.), 3.25 (s, 3H, OCH₃), 2.03 (s, 3H, CH₃, Ac.).

Anal. Calcd for C₆₆H₇₂O₁₄: C,72.77; H,6.66. Found: C,72.89; H,6.82.

Methyl 2,3,4-tri-O-benzyl-6-O-(3,4,6,7-tetra-O-benzyl-L-glycero- α -D-manno-heptopyranosyl)-L-glycero- α -D-manno-heptopyranoside (21). To a solution of compound 20 (46 mg, 0.04mmol) in methanol (2.5 mL) was added potassium *tert*. butoxide. After stirring for 2 h, the solution was neutralized with Dowex W50 (H⁺-form) and the filtrate was concentrated under reduced pressure. The oil thus obtained was applied on a column of silica gel, and the column was eluted with CH₂Cl₂ followed by CH₂Cl₂/acetone (99/1, v/v). Concentration of the appropriate fractions gave 21 (40 mg, 0.038 mmol) as an oil. Rf 0.3 (acetone/CH₂Cl₂, 3/97, v/v), [α] +26.1° (c 1, CHCl₃). ¹H NMR (CDCl₃): δ 7.32-7.21 (35H, H-arom.), 4.84-3.87 (H-1 - H-7a,b, H-1' - H-7'a,b, 7xCH₂, Bn.), 3.27 (s, 3H, OCH₃).

Anal. Calcd for C₆₄ H₇₀O₁₃; C,73.40; H,6.74. Found: C.74.38, H,6.71.

Methyl 6-O-(L-glycero- α -D-manno-heptoyranosyl)-L-glycero- α -D-manno-heptopyranoside (22). Dimer 21 (40 mg, 0.038 mmql) was dissolved in methanol (5 mL) and hydrogenolysis was performed on palladium on active carbon. After stirring for 48 h, the catalyst was removed by filtration and washed with water. The combined filtrates were concentrated under reduced pressure to give 22 (10 mg, 63%) as an amorphous material. Rf 0.1 (methanol/ethyl acetate/water, 3/5/2, v/v/v), [α] +45.7° (c 1, water). ¹H NMR (D₂O): δ 5.16 (d, 1H, H-1, J₁₂=1.8 Hz), 4.74 (d, 1H, H-1, J₁₂=1.9 Hz), 4.12 (dt, 1H, H-6, J_{6.7b}=5.8 Hz, J₅₆=1.0 Hz), 4.02 (ddd, 1H, H-6', J_{5'b}=1.3 Hz, J_{6'7b}=5.8 Hz, J_{6'7b}=7.1 Hz), 3.98 (dd, 1H, H-2, J₂₃=3.0 Hz), 3.91 (dd, 1H, H-2', J₂₂₃=3.7 Hz), 3.90-3.7- (10H, H-3, H-4, H-5, H7a,b, H3', H4', H-5', H7'a,b); ¹³C NMR (D₂O): δ 102.0 (C- 99.3 (C-1'), 73.7, 73.4, 72.0, 71.9, 71.4, 71.2, 70.7, (C-2, C-3, C-4, C-5, C-6, C-2', C-3', C-4', C-5', C-6') 69.9, 69.7 (C-7, C-7'), 55.8 (OCH₃).
 Anal. Calcd for C₁₅H₂₈O₁₃: C,43.27; H,6.78. Found: C,43.18; H6.70.

ACKNOWLEDGEMENT

Financial support for this work was generously provided by the World Health Organisation. We wish to thank Mr A.W.M. Lefeber and Dr. C. Erkelens for recording the ¹H and ¹³C NMR spectra.

REFERENCES AND NOTES

- L. Kenne and B. Lindberg in The Polysaccharides, G.O. Aspinall (Ed.), Vol. 2, 287-363, Academic Press, New York, (1983).
 H. Brade and E.Th. Rietschel, Eur. J. Biochem., 145, 231 (1984).
 H. Brade, U. Zahringer, E. Th. Rietschel, R. Christian, G. Schulz and F. Unger, Carbohydr. Res., 134, 157, (1984).
- H. Brade and C. Galanos, Infect. Immun., 42, 250, (1983).
 O. Luderitz. A.M. Staub and O. Westphal, Bacteriol. Rev. 30, 192 (1966).
 A.D. Elibein and C. Heath, J. Biol. Chem., 240, 1919, (1965).
- 3. G.J.P.H. Boons, P.A.M. van der Klein, G.A. van der Marel and J.H. van Boom, *Recl. Trav. Chim. Pays Bas*, 107, 507 (1988).
- 4. G.J.P.H. Boons, G.A. van der Marel and J.H. van Boom, *Tetrahedron Lett.* 30, 229 (1989).
- G.J.P.H. Boons, P.A.M. van der Klein, G.A. van der Marel and J.H. van Boom, Recl. Trav. Chim. Pays Bas, 109, 255 (1990).
 P.A.M. van der Klein, G.J.P.H. van Boons, G.H. Veeneman, G.A. van der Marel and J.H. van Boom, Tetrahedron Lett., 30, 5477 (1989).
- J.H. Banoub, Yeun-Min Choy, F. Michon and D.H. Shaw, Carbohydr. Res., 114, 265 (1983).
 J.H. Banoub, F. Michon and D.H. Shaw and *ibid.*, 128, 203 (1984).
 J.H. Banoub, F. Michon and D.H. Shaw, Can. J. Biochem. Cell Biol., 63, 1199 (1985).
- 7. H. Paulsen, A. Wulff and A.C. Heitmann, Liebigs Ann. Chem., 1073 (1988).
- 8. K. Tamao and N. Yshida, Tetrahedron. Lett., 25, 4245 (1984).
- 9. G.J.P.H. Boons, G.A. van der Marel and J.H Boom, Recl. Trav. Chim. Pays Bas, 108, 339 (1989).
- 10. P.J. Garegg, S. Oscarson and M. Szonyi, Carbohydr. Res., 205, 125 (1990).

- G.J.P.H. Boons, M. Overhand, G.A. van der Marel and J.H. van Boom, Carbohydr. Res., 192, c1-c4, (1989).
 G.J.P.H. Boons, M. Overhand, G.A. van der Marel and J.H. van Boom, Angew. Chem. Int. Ed. Engl., 28, 1504 (1989).
- 12. I. Fleming and P.E.J. Sanderson, Tetrahedron Lett., 28, 4429 (1987).
- 13. S. Czernecki, G. Georgoulis and C. Provelenghiou, Tetrahedron Lett., 3535 (1976).
- 14. D. Peterson, J. Org. Chem. 33, 780 (1968).
- H. Paulsen and A.C. Heitman, *Liebigs Ann. Chem.*, 1061 (1988).
 G.J.P.H. Boons, M. Overhand, G.A. van der Marel and J.H. van Boom, *Carbohydr. Res.*, 192, c1-c4, (1989).
- 16. H. Paulsen and O. Lockhoff, Chem. Ber., 111, 2334 and 2348 (1978),
- 17. M. Kloosterman, M.P. de Nijs and J.H. van Boom, J. Carbohydr. Chem. 5, 215 (1986).
- B. Helferich and W. Olst, Chem. Ber. 95, 2612 (1962).
 H.M. Flowers, Methods Carbohydr. Chem. 6, 474 (1972).
- 19. W. Kaca, J. de Jong-Leuvenink, U. Zahringer, E. Th. Rietschel, H. Brade, J. Verhoef and V. Sinnwell, *Carbohydr. Res.*, 179, 289 (1988).
- 20. S. Hanessian and J. Banoub, Carbohydr. Res., 53, C13-C16 (1977).
- 21. (Phenyldimethylsilyl)methyl chloride is commercially available from Aldrich chemical company.
- 22. An alternative approach in which a terminal olefinic precursor of LD-Hepp, prepared by stereospecific condensation of a similarly protected aldehyde 3 with vinyl magnesium bromide, plays a central role-was published recently [see: Y. Chaplear et al., J. Chem. Soc. Perkin Trans I, 3091 (1990)]. However, it has to be seen whether the presence of a terminal olefinic function in the above LD-Hepp precursor is compatible with an allyl protecting group strategy.